

Optimizing RSV Treatment: Insights into Targeted Drug Delivery Strategies and Public Health Implications

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ABSTRACT

Respiratory Syncytial Virus (RSV) remains a pressing health issue worldwide, with infants, young children, and elderly individuals, especially vulnerable. In the present study, we reviewed available treatment modalities, Their efficacy rates were measured, and possible improvements of both therapeutic methods and pharmacological delivery methods were explored. Through analysis of recent literature and data, we evaluated the current efficacy of bronchodilators, anti-inflammatory agents, and monoclonal antibodies work in RSV treatment. Notably, while bronchodilators and corticosteroids only offer monoclonal antibodies, such as Palivizumab, confer preventive benefits among those at high risk but with limited alleviation. Additionally, emerging therapeutic options—such as RNA interference and nucleoside analogs—have demonstrated initial potential in thereby minimizing RSV replication. These findings emphasize the need for targeted therapy that is not only specific but also effective in reducing the complications and mortality burden caused by RSV. Others, like Palivizumab, are showing prospects for high-risk groups, RNA interference treatments may offer a new strategy in directly targeting viral replication. Perhaps the best route to reduction incorporates both prevention and direct intervention in a multifaceted approach. Development of RSV, and further research into siRNA-based therapies with enhanced drug delivery. Methodological approaches are important, as our findings have pointed out, for protecting the most vulnerable from RSV.

Introduction

Further supporting the fact that RSV is one of the leading causes of child morbidity and mortality worldwide are estimates for 2015 that it accounted for 33.1 million cases of acute respiratory infections, translating into 3.2 million hospitalizations and nearly 60,000 in-hospital deaths. The true burden of RSV infection is likely many times higher than this, as most cases in developing countries go unreported due to limited access to health facilities. RSV remains one of the most leading causes of severe illnesses and deaths among under-five-year-old children, most of whom are admitted to hospitals due to diseases such as bronchiolitis and pneumonia. However, in the last few years, awareness has been growing about the severity of infection in the aged, whose immune system is very weak. These patterns illustrate the need for international research in understanding its regional impacts and thus adapting the treatment strategies to fit the resources of local healthcare systems.

In the United States, for example, several researchers have explored the complications involved in the RSV treatment process, together with problems of active drug delivery. Valuable studies, conducted by Falsey et al. in 2005 among others, have indeed provided valuable information about the impact RSV has on high-risk adults and have thus shifted the paradigm. These findings from these studies further illustrate the differential burden of the virus between children and adults, pointing to the challenges of offering care to populations with different levels of service access. Such gaps in inequity, if addressed, would become important in the protection of the most vulnerable people.

In Pennsylvania, individual studies have highlighted specific challenges that healthcare service providers encounter in the handling of RSV. Numerous research into outbreaks in care homes and within children suffering from fragile body immunity has contributed to the achievement of workable means through which to address such regional issues. Research by Balasubramani et al. (2022), conducted specifically in Allegheny County, has also identified the strain that RSV places on hospitals, especially among adults. This thus calls for some area-based studies, proper monitoring, and specific measures; all in a bid to formulating plans that actually respond to community-specific healthcare needs concerning the disease.

The study is important because it aims to fill critical gaps in current understanding with respect to the improvement of drug delivery systems in treating RSV infection. This research is a starting point for treatments that will eventually be not only effective but accessible to those who need them most, by tackling key challenges and proposing feasible solutions. Drawing from the conclusion of various studies, this work has described the difficulties in addressing RSV and pointed out the ways forward to more specific interventions. It thus has the potential to alter the course of therapeutic approaches and, consequently, reduce the global burden of RSV and offer, in the end, better services to the ones most affected.

Methodology

This study of RSV therapeutic interventions has placed much emphasis on the systems for administering drugs. The study formed the beginning in an extensive review of several sources; some of the sources reviewed were peer-reviewed articles, conference proceedings, and other literatures of repute for contributions from varied contributions so as to be able to have a broad understanding of the various available treatment options and also the different methodologies applicable to delivering drugs.

The study also entailed a critical review of different existing literature based on strict selection to ensure that only the highly valid and relevant studies were selected. Only credible and relevant sources were included to ensure that it would serve effectively toward achieving results that contribute to improvements in the treatment of RSV. Searches were made through systematic searches in reputed databases like PubMed and Google Scholar using very specific keywords to identify studies that were most applicable.

The clinical trial data concerned with both patient characteristics and treatment responses were reviewed on the effectiveness of their approaches. By such an analysis, one could realize the trend in RSV management: from antiviral drugs and monoclonal antibodies to new drug delivery systems. The findings testified which treatments were most promising toward improved care for the patients.

Ultimately, this study investigated recent drug delivery development. The study reviewed ongoing clinical trials in the attempt to obtain new therapeutic options and identify further improvements needed for the existing methodologies. This managed to further bring into light considerable knowledge by keeping up to date with the latest developments in the field that sheds light on routes to better therapies and improvements of the existing modalities. This approach aimed to achieve an overall understanding of the RSV treatment modalities by collating data from many sources. The aim of the study was to offer workable suggestions which could lead to the betterment of the patient conditions and reduce the extensive effects caused by RSV.

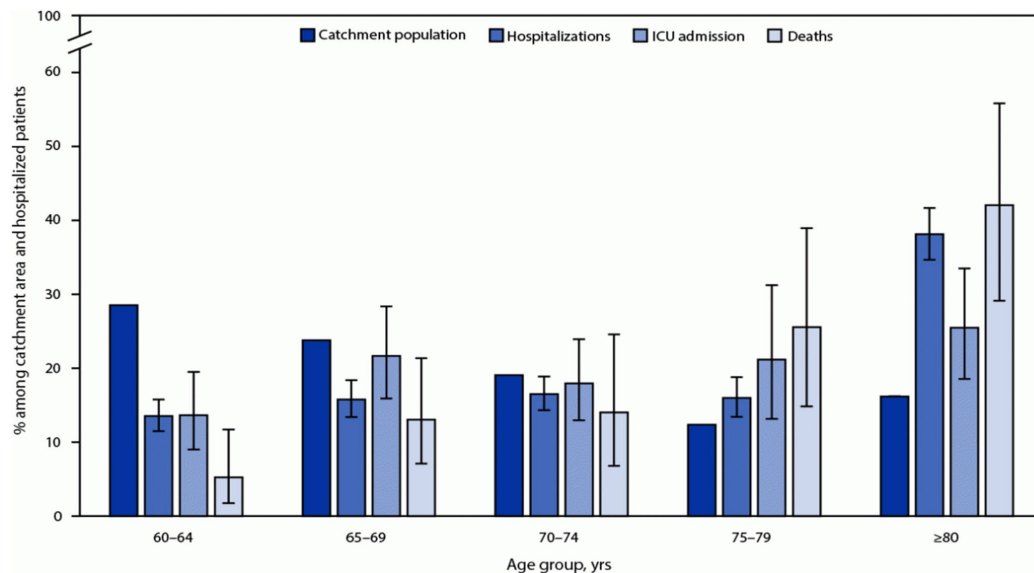


Figure 1. Distribution by Age Group of the Hospitalized Adults (Havers et al., 2023). Age distribution among persons aged ≥ 60 years residing in the surveillance network catchment area† and among laboratory-confirmed respiratory syncytial virus–associated hospitalizations, intensive care unit admissions, and in-hospital deaths — Respiratory Syncytial Virus–Associated Hospitalization Surveillance Network, 12 states, October 2022–April 2023.

Pathogenesis of RSV

In this regard, epidemiological studies have extensively identified select groups of infants who have a high risk of serious outcomes related to respiratory syncytial virus infection. However, approximately a quarter of the children who develop RSV lower respiratory tract infection requiring hospitalization are previously healthy with no coexisting health conditions and do not present recognizable risk factors for severe illness (García et al., 2010; Kristensen et al., 2012). Of these, 10-20% of the hospitalized infants will suddenly deteriorate and necessitate ICU admissions. Although the disease burden is remarkably huge, effective vaccines or targeted treatments have remained a mirage due to a gap in our understanding of the immune response to RSV and its relationship with disease severity.

Empirical research studies and clinical observations have both revealed marked heterogeneity in the severity of RSV infection among infected children (Hall et al., 2009). Predicting which patients will progress to worsening illness requiring hospitalization or ICU care based on physical examination and available diagnostic studies is particularly challenging. Therefore, attempts to identify clinical risk factors or biomarkers, such as cytokines, for determining the severity of RSV infection have been inconclusive so far. Both viral and host immune factors are believed to be important in the pathogenesis of RSV disease severity (DeVincenzo et al., 2005; Houben et al., 2010; Larrañaga et al., 2009; Sheeran et al., 1999; Bennett et al., 2007).

Role of Viral Loads

Several investigations have tried to correlate RSV loads measured in the respiratory tract either by culture or by real-time (RT)-PCR with disease severity and have provided conflicting results (Houben et al., 2010; Sheeran et al., 1999; Saleeby et al., 2011; Martin et al., 2008; Wright et al., 2002). Many of these studies often included patients of different ages and stages of infection, which could have obscured analyses due to confounding variables. A recent prospective study, therefore, sought to address this by evaluating 1764 children less than two years of age who were admitted to the hospital due to RSV LRTIs and discovered that the higher the genomic RSV load, the greater the chances of

children requiring PICU admission and having longer hospitalization, even after adjustment for other confounding factors (Hasegawa et al., 2014).

Researchers have also looked into how various subtypes of RSV, like types A and B, and their genotypes, might affect the severity of the disease. While some studies identified a significant correlation between RSV type A and disease severity, others failed to replicate these findings (Jafri et al., 2013). Nevertheless, during typical RSV outbreaks, which usually occur annually from late fall through early spring in temperate climates, only a limited number of RSV strains tend to circulate. Consequently, it remains unclear why previously healthy children infected with the same viral strain exhibit such diverse clinical manifestations, suggesting that an abnormal host immune response or immune "insufficiency" may significantly contribute to the variability in disease phenotypes.

Host Immune Response

Until recently, it was postulated that severe RSV infection was associated with an exaggerated inflammatory response. Others have shown that host innate immune responses are actually inadequately activated or even suppressed in infants with severe RSV disease (Houben et al., 2010; Sheeran et al., 1999; Mehta et al., 2014; Mella et al., 2012). These observations represent a paradigm shift and suggest that weak, rather than intact or robust innate immune responses are associated with enhanced acute disease severity and may contribute to the chronic/ persistent airway disease observed in a subset of children after RSV LRTI (as shown in Fig. 1). Innate immune functional responses were recently categorized in a cohort of previously healthy infants hospitalized with a first episode of RSV LRTI. Unstimulated plasma innate immunity cytokines including IL-6, IL-8, and also IL-10 were modestly increased in infants RSV LRTI compared with healthy controls, and this was independent of the severity of the disease. On the other hand, children with RSV LRTI requiring ICU treatment had significantly lower production capacity of TNF- α after LPS stimulation compared with infants with less severe RSV bronchiolitis hospitalized in the inpatient floor, and with healthy controls. Lower TNF- α production capacity independently predicted longer duration of hospitalization after adjusting for age, gender, days of symptoms at enrollment, presence of fever, and RSV loads (Mehta et al., 2014).

Using a completely different and unbiased methodology, genome-wide transcriptional profiling, studies suggest the existence of an impaired immune response in the more severe forms of RSV LRTI, and emphasizes the value of blood host gene signatures as a novel tool to better understand the pathogenesis of RSV infection (Bucasas et al., 2013; Ioannidis et al., 2012). These results suggest that children with severe RSV infection exhibit defective innate immune responses. Whether these children are born with an already impaired innate immune response, and RSV just uncovers their abnormal immune system, or whether it is just RSV by itself that causes such a profound suppression of the immune response it is unknown.

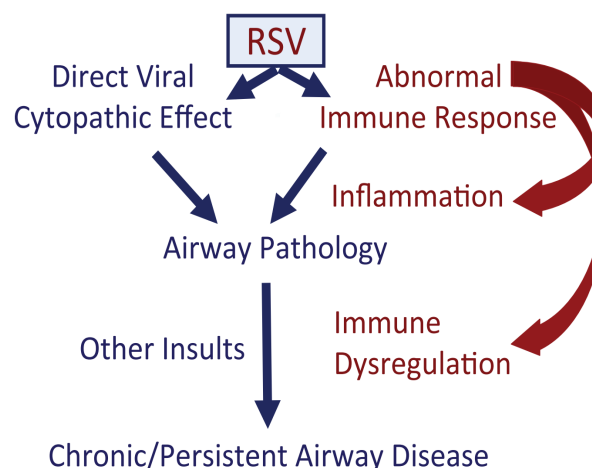


Figure 2. Pathogenesis of RSV bronchiolitis (Mejias, 2015). A complex interaction between viral factors and the host immune response contributes to the severity of RSV disease. Recent studies have challenged the conventional view that an exaggerated innate immune response is solely responsible for disease severity. Instead, findings suggest that impaired immune responses, along with other factors like environmental influences and airway size, are associated with increased acute disease severity. These factors may also contribute to the development of chronic or persistent airway disease observed in some children following RSV LRTI.

Current Treatments

Bronchodilators

Current standard therapy for RSV bronchiolitis in small children, especially those presenting with severe respiratory distress, includes nebulized albuterol, epinephrine, and heliox. In this setting, bronchodilators play their main role through symptomatic improvement by bronchial dilatation, decreasing wheezing and airway obstruction. Nevertheless, the wide usage of bronchodilators in the clinical setting remains without clear and proven long-term benefits to the actual improvement of the patient. Some studies indicate that bronchodilators may confer transient benefits in severe bronchiolitis; however there are few effective therapies for the condition because they appear ineffective for the more critical endpoints of length of stays, need for supplemental oxygen, or rates of admission to an ICU (Sugawara et al., 2002; Cane et al., 1994).

In general, the role of these drugs in the management of RSV bronchiolitis is still in doubt, despite their widespread usage. Administering them is routine, but again, the real benefit is not considered, with big variations in response for each child. Researchers look towards the subgroup of patients that would particularly benefit from them, like patients with certain conditions or with severer symptoms. Meanwhile, current research is focused on the search for new delivery modes and preparations to ensure the therapies are safer and more efficient. This all will certainly help clinicians give more authoritative advice on when and how to use or avoid bronchodilators, thus raising the standard of care for the children with RSV bronchiolitis.

Anti-Inflammatory Medications

A number of studies revealed the low efficacy of nebulized and systemic corticosteroids in acute RSV bronchiolitis both as an independent agent and in combination with bronchodilators (Wright et al., 2002; Crowe, 2001; Staat, 2002; Openshaw, 2002). Intravenous dexamethasone administration, on the other hand, resulted in a prolonged viral shedding that diminishes its use in treating RSV infection (DeVincenzo et al., 2008; Sugawara et al., 2002). Similarly, other class of anti-inflammatory agent tested for therapeutic effect on bronchiolitis, that of leukotriene antagonists showed surprising ineffectiveness, underlining again the practical uselessness of anti-inflammatory approach (Cane et al., 1994)

However, the balance to this growing evidence of the futility of anti-inflammatory treatments for RSV bronchiolitis is an overwhelming pressure on early intervention on symptomatic treatment of the disease; hence, the majority of the patients go through corticosteroids and other anti-inflammatory agents despite the lack of evidence in controlled trials and meta-analyses showing clinically important benefits of these interventions. The clinical practice-research finding gap is a pointer to the fact that treatment decisions have to be facilitated by the latest available evidence-based guidelines in order to ensure that health care provided is adequate but not excessive.

Prophylaxis and Vaccines

Currently, palivizumab is the only available prophylaxis against respiratory syncytial virus infection and provides protection against severe RSV disease. This humanized monoclonal antibody is directed against a well-conserved epitope of the RSV F fusion protein, and protection is maintained by monthly injections during seasonal outbreaks of RSV (Griffiths et al., 2017). Palivizumab is highly effective in preventing severe RSV disease, mainly in premature infants and children with certain types of cardiopulmonary disease. However, the clinical use of this is limited to prophylaxis only, because the antibody does not offer any benefit as far as treatment of active infection is concerned (Meissner, 2016). Thus, there is an urgent need for therapies against RSV infection, and that constitutes a great need for continued research and development activity.

Despite the urgent need for an RSV vaccine, clinical development had been at a standstill due to historical obstacles, one of which was the 1960 controlled clinical trial that resulted in enhanced RSV disease and fatalities upon the administration of a formalin-inactivated RSV virus (Mazur et al., 2017). This resulted in a much more cautious and thoughtful approach being proposed for the development of RSV vaccines, taking into consideration the delicate balance between the pathogenicity and the immunogenic properties of the vaccine strains Meissner, 2016. The quest to achieve a safe and effective RSV vaccine continued unabated, as many obstacles have been overcome by the investigators in developing a perfectly constructed vaccine and ascertaining its use.

Licensed Antivirals

Ribavirin

Ribavirin is a guanosine analog and an antiviral agent known for its broad spectrum of activity against RNA viruses, including RSV. Early clinical trials, including small-scale, double-blind, placebo-controlled trials indicated that aerosolized ribavirin could be used as a treatment given its administration during the first days of illness in infants with RSV infection. However, benefit and risk issues of overall concern have made its use limited. Consequently, the American Academy of Pediatrics recommends that ribavirin should not be used routinely in children. Ribavirin use is recommended only in very carefully selected cases, especially in the treatment of severe RSV diseases in those at high-risk (American Academy of Pediatrics, 2012). This recommendation highlights the consideration that ribavirin should be used when the patient has no other alternatives.

Monoclonal Antibodies

The introduction of monoclonal antibodies significantly transformed RSV treatment options. In the 1980s, an intravenous polyclonal immunoglobulin preparation (RSV-IVIG) containing high levels of neutralizing antibodies against RSV demonstrated its ability to reduce hospitalizations caused by severe RSV infections in high-risk children. The findings from the PREVENT study confirmed that RSV-IVIG could effectively prevent RSV-related hospitalizations in high-risk infants, including those born prematurely and those with bronchopulmonary dysplasia (The PREVENT Study Group, 1997). This laid the groundwork for future antibody-based therapies aimed at preventing severe RSV complications.

Novel Treatments

RNA Interference

Among the new therapeutic approaches to RSV, one of the most novel approaches is the use of siRNAs that possess specific anti-RSV activity (Bitko et al., 2004). These siRNAs are double-stranded segments of RNA that inherently function to control protein synthesis through targeting of post-transcriptional mRNA. Researchers are thereby capable of artificially targeting and degrading RSV mRNA through the use of specifically designed siRNAs for therapeutic purposes (DeVincenzo et al., 2010). ALN-RSV01 is the first RNA interference therapeutic to be brought into clinical use and is developed to target the mRNA encoding the nucleocapsid protein, simply known as the N-protein, of RSV. By binding and consequently cleaving the mRNA of RSV encoding the N-protein, ALN-RSV01 effectively blocks the synthesis of the N-protein. Experimental studies have proved that ALN-RSV01 possesses anti-viral activity against RSV subtype A and RSV subtype B. DeVincenzo et al., 2008; DeVincenzo et al., 2010.

In 2006, Alnylam successfully completed two phase I trials with healthy male adults. One year later, it completed a phase II trial with healthy adults inoculated with RSV. Aerosolized ALN-RSV01 had a statistically significant antiviral effect, including lower infection rates post-inoculation, and, in cases of infection, reduced symptom severity scores, as obtained from, DeVincenzo et al., 2010. Between 2009 and 2012, two phase II studies of ALN-RSV01 in lung transplant patients with acute RSV infection were completed. Although there was no statistically significant antiviral effect, ALN-RSV01 was well tolerated and not associated with treatment-emergent immunogenicity. Importantly, it enhanced the daily cumulative symptom scores and clearly mitigated the occurrence of new onset/worsening bronchiolitis obliterans syndrome, a disease similar to organ rejection post-transplantation (Zamora et al., 2011).

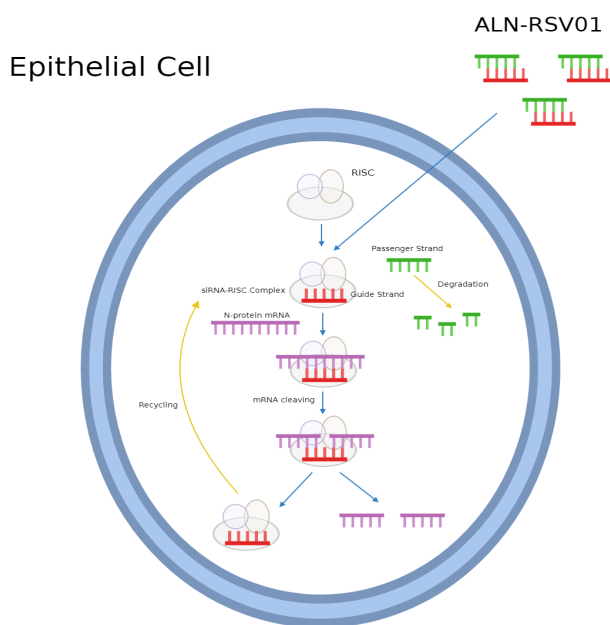


Figure 3. ALN-RSV01 mechanism of action (Created in BioRender by Lakkimsetti, 2024). The RNA-induced silencing complex (RISC) plays a key role in RNA interference (RNAi), a process essential for gene silencing and viral defense. Upon administration, the two strands of ALN-RSV01 separate, with the guide strand integrating into the RISC. This integration enables the complex to recognize complementary mRNA sequences using single-stranded RNA fragments, such as microRNA (miRNA), or double-stranded small interfering RNA (siRNA). Once the N-protein mRNA is identified, it is cleaved by the RISC, leading to the release of mRNA fragments. The guide strand directs the siRNA-RISC complex, while the passenger strand is degraded. The complex then recycles itself to repeat the process, ensuring continuous gene silencing and viral inhibition.

Nucleoside Analogs

Nucleoside analogs represent one of the main classes of antiviral drugs that, by structure, feature similarity with natural nucleosides and, therefore, easily interfere with viral replication processes. These artificial compounds, once integrated into either viral RNA or DNA, interfere with proper synthesis and further viral growth. Such is the compound known as ALS-008176, serving as a prodrug for ALS-008112-a strong inhibitor of RSV polymerase where ALS-008112 is a cytidine nucleoside analog of high oral bioavailability. It acts through its active metabolite, ALS-8112-TP, upon entrance into the respiratory tract. Inhibiting RNA synthesis by competing with CMP, this molecule results in premature chain termination and, therefore, the blockade of the replicative cycle of RSV (DeVincenzo et al., 2015). Of note, ALS-8112-TP itself does not inhibit human RNA or DNA polymerases, further underlining its selectivity and safety profile (Wang et al., 2015).

Between 2013 and 2015, Alios BioPharma, in cooperation with Janssen, had successfully conducted three phase I studies aimed at testing drug safety in healthy adult subjects. In 2014, results of a phase II viral challenge study conducted in adults evidenced potent antiviral activity of the compound, with symptom improvement for RSV. By 2016, it was in a phase IIa trial, assessing efficacy in RSV infections in adults. The compound, now called JNJ-64041575, continued into 2018 in the clinic. Janssen finished a phase I study of the antibody in infants up to 12 months of age who had been hospitalized due to RSV bronchiolitis. It began two phase II studies: one in children below 36 months of age and one in adults. However, this trial has also been put on hold, pending further protocol modifications and data analyses (DeVincenzo et al., 2015).

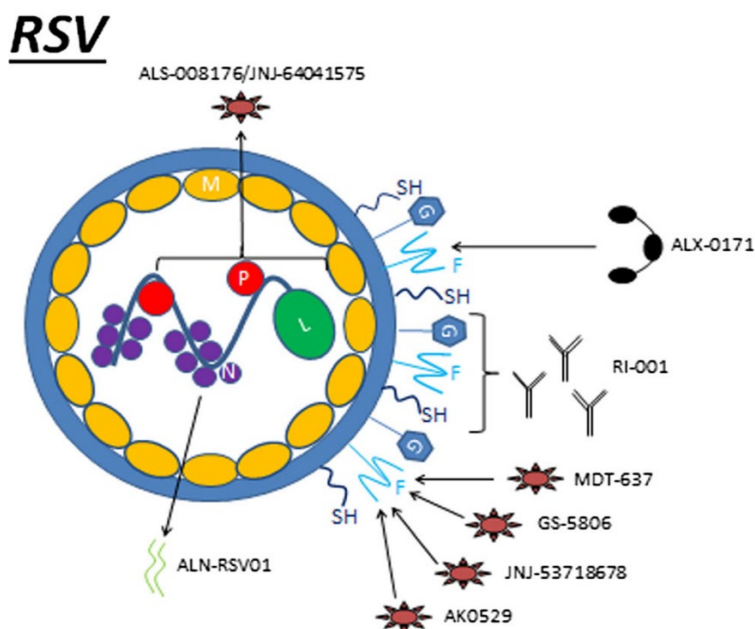


Figure 4. RSV targets and antivirals (Xing & Proesmans, 2019). The arrows illustrate the specific RSV proteins targeted by different antivirals. ALS-008176/JNJ-64041575 is designed to target the entire polymerase complex, including the P, N, and L proteins, whereas ALN-RSV01 is a siRNA that specifically targets N-mRNA. RI-001 is a polyclonal antibody that recognizes various RSV surface epitopes. The F protein is a common target for several antibodies, such as ALX-0171, and antivirals, including MDT-637, GS-5806, JNJ53718678, and AK0529.

Results and Discussion

This research aimed to assess the effectiveness of current RSV treatments, specifically focusing on bronchodilators, anti-inflammatory drugs, and monoclonal antibodies. Though nebulized albuterol, epinephrine, and heliox are commonly in use, being bronchodilators, they offered only scanty transient gain in the respiratory variables. The same gains did not always translate into good long-term outcomes of the disease, such as shortened stays in hospitals or ICUs. This realization has curtailed the routine administration of bronchodilators in the treatment protocols for RSV. Anti-inflammatory drugs, particularly systemic and nebulized corticosteroids, are also of limited value in managing RSV bronchiolitis. Corticosteroids have no significant effect on important clinical outcomes related to duration of mechanical ventilation, length of stay, and inflammation markers.

Additionally, their use was linked to prolonged viral shedding hence raising other questions concerning their mode of action in the management of RSV. Monoclonal antibodies especially Palivizumab remain the best prophylaxis. Palivizumab works by reduction of the rate of hospitalisation in high-risk infants without any activity against acute RSV infection. Given the high cost and limited scope of Palivizumab, it is recommended only for specific high-risk groups, such as premature infants and those with cardiopulmonary disorders. Emerging treatments such as RNA interference (siRNAs) and nucleoside analogs, including ALS-008176, show promise. Therapies such as ALN-RSV01 involving siRNA against RSV N-protein mRNA have already been shown to possess anti-viral activity by symptom severity reduction in clinical trials. Certain nucleoside analogs are promising, and for example, one such analog ALS-008112 selectively inhibits the RSV polymerase. While treatment of RSV is mainly these, further study into long-term efficacy and safety will be required.

Conclusion

The therapeutic realm of RSV is complex, and there is much to be learned about the understanding and management of this infection. Despite efforts at determining risk groups, many children hospitalized with severe RSV infection have no antecedent complicating illness that might aid predictive models. Currently available diagnostic tools and therapeutic interventions often do not meet clinical needs, pointing to a great need for further research on both viral and host immune responses that modulate disease severity. The use of bronchodilators is also very common but offers very variable benefits; its role in the treatment of RSV is still debated. Similarly, anti-inflammatory agents demonstrate limited efficacy, often with prolongation of viral shedding unaccompanied by significant clinical improvement. These attributes tend to explain the dire need for evidence-based policies to advocate for the effective treatment methods and forbid the unnecessary procedures. Since Palivizumab is the only licensed prophylaxis, its application is mainly on prevention, applied only for high-risk individuals due to its high cost and small scope of application.

This therefore forms the rationale for developing new therapeutic interventions. Newer therapies, namely RNA interference agents, including ALN-RSV01, are promising and have shown anti-viral activity with symptom reduction in clinical trials but need further validation and optimization. Vaccine development against RSV remains an elusive task because an unusually delicate balance exists between immunogenicity and safety, complicated by past disappointments. Further technological advances in genetic profiling and immunological research may open newer frontiers to be exploited in understanding and fighting against RSV. Finally, the current status of treatment against RSV urges the need for therapies with higher efficacies. The drugs like bronchodilators and corticosteroids hold limited clinical benefits, and their routine use needs revision.

Monoclonal antibodies such as Palivizumab constitute a good prophylaxis; however, its high cost and limited application call for interest in other modes of prevention. Newer therapeutic therapies such as siRNAs and nucleoside analogs have a great promise toward targeting RSV at a molecular level, hence a potentially promising avenue of study. Further studies need to be aimed at optimization of these therapies, investigation of other available anti-viral drugs, and modes of drug delivery for improving the above outcomes. Moreover, continued investigation into the

immune response to RSV is critical to developing more targeted and effective interventions, potentially alleviating the global burden of RSV. Future studies should prioritize these areas, as they present key opportunities for advancing treatment strategies and mitigating the impact of RSV, particularly in vulnerable populations like infants and children.

Limitations

This study focuses exclusively on RSV treatments and their efficacy, with particular attention to emerging therapeutic interventions. One limitation was the reliance on secondary data sources, which may introduce bias in the findings. Additionally, clinical trials and research studies were not uniformly accessible, limiting the comprehensiveness of the analysis. Some difficulties encountered include the complex nature of synthesizing data from various studies, as well as challenges in gathering up-to-date information on ongoing clinical trials. Future research could expand the scope to include a broader analysis of RSV prevention methods or explore treatments for other respiratory viruses.

Acknowledgments

My deepest appreciation to Professor Kathryn Wilwohl and Professor Virgel Torremocha, for their guidance, insightful advice, and invaluable input to improve the content of my paper. My acknowledgement also goes to the Gifted Gabber team for enriching my academic journey through this research program. Above all, my heartfelt thanks to my family for encouraging me on my scientific research quest.

References

- Agius, G., Dindinaud, G., Biggar, R. J., Peyre, R., Vaillant, V., Ranger, S., Poupet, J. Y., Cissé, M., & Castets, M. (1990). An epidemic of respiratory syncytial virus in elderly people: Clinical and serological findings. *Journal of Medical Virology*, 30(2), 117–127. <https://doi.org/10.1002/jmv.1890300208>
- Balasubramani, G., Nowalk, M. P., Eng, H., & Zimmerman, R. K. (2022). Estimating the burden of adult hospitalized RSV infection using local and state data - methodology. *Human Vaccines & Immunotherapeutics*, 18(1). <https://doi.org/10.1080/21645515.2021.1958610>
- Bennett, B. L., Garofalo, R. P., Cron, S. G., Hosakote, Y. M., Atmar, R. L., Macias, C. G., & Piedra, P. A. (2007). Immunopathogenesis of respiratory syncytial virus bronchiolitis. *The Journal of Infectious Diseases*, 195(10), 1532–1540. <https://doi.org/10.1086/515575>
- Bitko, V., Musiyenko, A., Shulyayeva, O., & Barik, S. (2004). Inhibition of respiratory viruses by nasally administered siRNA. *Nature Medicine*, 11(1), 50–55. <https://doi.org/10.1038/nm1164>
- Bucasas, K. L., Mian, A., Demmler-Harrison, G. J., Caviness, A. C., Piedra, P. A., Franco, L. M., Shaw, C. A., Zhai, Y., Wang, X., Bray, M. S., Couch, R. B., & Belmont, J. W. (2013). Global gene expression profiling in infants with acute respiratory syncytial virus bronchiolitis demonstrates systemic activation of interferon signaling networks. *The Pediatric Infectious Disease Journal*, 32(2), e68–e76. <https://doi.org/10.1097/inf.0b013e318278b4b3>
- Cane, P. A., Matthews, D. A., & Pringle, C. R. (1994). Analysis of respiratory syncytial virus strain variation in successive epidemics in one city. *Journal of Clinical Microbiology*, 32(1), 1–4. <https://doi.org/10.1128/jcm.32.1.1-4.1994>
- Chanock, R. M., Roizman, B., & Myers, R. (1957). Recovery from Infants with Respiratory Illness of a Virus Related to Chimpanzee Coryza Agent (CCA). *American Journal of Epidemiology*, 66(3), 281–290. <https://doi.org/10.1093/oxfordjournals.aje.a119901>
- Crowe, J. E. (2001). Influence of Maternal Antibodies on Neonatal Immunization against Respiratory Viruses. *Clinical Infectious Diseases*, 33(10), 1720–1727. <https://doi.org/10.1086/322971>

- DeVincenzo, J. P., Cehelsky, J., Alvarez, R., Elbashir, S. M., Harborth, J., Toudjarska, I., Nechev, L., Murugaiah, V., Van Vliet, A., Vaishnav, A., & Meyers, R. (2008). Evaluation of the safety, tolerability and pharmacokinetics of ALN-RSV01, a novel RNAi antiviral therapeutic directed against respiratory syncytial virus (RSV). *Antiviral Research*, 77(3), 225–231. <https://doi.org/10.1016/j.antiviral.2007.11.009>
- DeVincenzo, J. P., Lambkin-Williams, R., Wilkinson, T., Cehelsky, J., Nochur, S. V., Walsh, E. E., Meyers, R., Gollob, J., & Vaishnav, A. (2010). A randomized, double-blind, placebo-controlled study of an RNAi-based therapy directed against respiratory syncytial virus. *Proceedings of the National Academy of Sciences of the United States of America*, 107(19), 8800–8805. <https://doi.org/10.1073/pnas.0912186107>
- DeVincenzo, J. P., Saleeby, C. M. E., & Bush, A. (2005). Respiratory syncytial virus load predicts disease severity in previously healthy infants. *The Journal of Infectious Diseases*, 191(11), 1861–1868. <https://doi.org/10.1086/430008>
- Falsey, A. R., Hennessey, P., Formica, M. A., Cox, C., & Walsh, E. E. (2005). Respiratory syncytial virus infection in Elderly and High-Risk adults. *The New England Journal of Medicine*, 352(17), 1749–1759. <https://doi.org/10.1056/nejmoa043951>
- Falsey, A. R., Hennessey, P., Formica, M. A., Cox, C., & Walsh, E. E. (2005). Respiratory syncytial virus infection in Elderly and High-Risk adults. *the New England Journal of Medicine (Print)*, 352(17), 1749–1759. <https://doi.org/10.1056/nejmoa043951>
- Falsey, A. R., Treanor, J. J., Betts, R. F., & Walsh, E. E. (1992). Viral respiratory infections in the Institutionalized elderly: Clinical and epidemiologic findings. *Journal of the American Geriatrics Society*, 40(2), 115–119. <https://doi.org/10.1111/j.1532-5415.1992.tb01929.x>
- García, C. G., Bhore, R., Soriano-Fallas, A., Trost, M., Chason, R. D., Ramilo, O., & Mejías, A. (2010). Risk factors in children hospitalized with RSV bronchiolitis versus Non-RSV bronchiolitis. *Pediatrics*, 126(6), e1453–e1460. <https://doi.org/10.1542/peds.2010-0507>
- García, C. S., Soriano-Fallas, A., Lozano, J., Leos, N. K., Gómez, A. M., Ramilo, O., & Mejías, A. (2012). Decreased innate immune cytokine responses correlate with disease severity in children with respiratory syncytial virus and human rhinovirus bronchiolitis. *The Pediatric Infectious Disease Journal*, 31(1), 86–89. <https://doi.org/10.1097/inf.0b013e31822dc8c1>
- Garofalo, R. P., Dorris, A., Ahlstedt, S., & Welliver, R. C. (1994). Peripheral blood eosinophil counts and eosinophil cationic protein content of respiratory secretions in bronchiolitis: relationship to severity of disease. *Pediatric Allergy and Immunology*, 5(2), 111–117. <https://doi.org/10.1111/j.1399-3038.1994.tb00227.x>
- Garofalo, R. P., Kimpen, J. L. L., Welliver, R. C., & Ogra, P. L. (1992). Eosinophil degranulation in the respiratory tract during naturally acquired respiratory syncytial virus infection. *The Journal of Pediatrics*, 120(1), 28–32. [https://doi.org/10.1016/s0022-3476\(05\)80592-x](https://doi.org/10.1016/s0022-3476(05)80592-x)
- Glezen, W. P. (1986). Risk of primary infection and reinfection with respiratory syncytial virus. *Archives of Pediatrics & Adolescent Medicine*, 140(6), 543. <https://doi.org/10.1001/archpedi.1986.02140200053026>
- Gonik, B. (2019). The burden of respiratory Syncytial virus infection in Adults and Reproductive-Aged women. *Global Health, Science and Practice*, 7(4), 515–520. <https://doi.org/10.9745/ghsp-d-19-00121>
- Griffiths, C., Drews, S. J., & Marchant, D. (2017a). Respiratory syncytial Virus: Infection, detection, and new options for prevention and treatment. *Clinical Microbiology Reviews*, 30(1), 277–319. <https://doi.org/10.1128/cmr.00010-16>
- Hall, C. B., Powell, K. R., MacDonald, N. E., Gala, C. L., Menegus, M. E., Suffin, S. C., & Cohen, H. J. (1986). Respiratory Syncytial Viral Infection in Children with Compromised Immune Function. *The New England Journal of Medicine*, 315(2), 77–81. <https://doi.org/10.1056/nejm198607103150201>
- Hall, C. B., Weinberg, G. A., Iwane, M. K., Blumkin, A., Edwards, K. M., Staat, M. A., Auinger, P., Griffin, M. R., Poehling, K. A., Erdman, D. D., Grijalva, C. G., Zhu, Y., & Szilagyi, P. G. (2009). The burden of

- respiratory syncytial virus infection in young children. *The New England Journal of Medicine*, 360(6), 588–598. <https://doi.org/10.1056/nejmoa0804877>
- Harrington, R. D., Hooton, T. M., Hackman, R. C., Storch, G. A., Osborne, B. M., Gleaves, C. A., Benson, A. M., & Meyers, J. D. (1992). An outbreak of respiratory syncytial virus in a bone marrow transplant center. *The Journal of Infectious Diseases*, 165(6), 987–993. <https://doi.org/10.1093/infdis/165.6.987>
- Hasegawa, K., Jartti, T., Mansbach, J. M., Laham, F. R., Jewell, A. M., Espinola, J. A., Piedra, P. A., & Camargo, C. A. (2014). Respiratory syncytial virus genomic load and disease severity among children hospitalized with bronchiolitis: multicenter cohort studies in the United States and Finland. *The Journal of Infectious Diseases*, 211(10), 1550–1559. <https://doi.org/10.1093/infdis/jiu658>
- Havers, F. P., Whitaker, M., Melgar, M., Chatwani, B., Chai, S. J., Alden, N. B., Meek, J., Openo, K. P., Ryan, P. A., Kim, S., Lynfield, R., Shaw, Y. P., Barney, G., Tesini, B. L., Sutton, M., Talbot, H. K., Olsen, K. P., Patton, M. E., Kirley, P. D., . . . Staten, H. (2023). Characteristics and Outcomes Among Adults Aged ≥60 Years Hospitalized with Laboratory-Confirmed Respiratory Syncytial Virus — RSV-NET, 12 States. *MMWR Morbidity and Mortality Weekly Report*, 72(40), 1075–1082. <https://doi.org/10.15585/mmwr.mm7240a1>
- Heilman, C. A. (1990). From the National Institute of Allergy and Infectious Diseases and the World Health Organization: Respiratory syncytial and Parainfluenza viruses. *Journal of Infectious Diseases*, 161(3), 402–406. <https://doi.org/10.1093/infdis/161.3.402>
- Houben, M. L., Coenjaerts, F. E. J., Rossen, J. W. A., Belderbos, M. E., Hofland, R. W., Kimpen, J. L. L., & Bont, L. (2010). Disease severity and viral load are correlated in infants with primary respiratory syncytial virus infection in the community. *Journal of Medical Virology*, 82(7), 1266–1271. <https://doi.org/10.1002/jmv.21771>
- Ioannidis, I., McNally, B. A., Willette, M., Peeples, M. E., Chaussabel, D., Durbin, J. E., Ramilo, O., Mejías, A., & Flaño, E. (2012). Plasticity and Virus Specificity of the Airway Epithelial Cell Immune Response during Respiratory Virus Infection. *Journal of Virology*, 86(10), 5422–5436. <https://doi.org/10.1128/jvi.06757-11>
- Jafri, H. S., Wu, X., Makari, D., & Henrickson, K. J. (2013). Distribution of respiratory syncytial virus subtypes A and B among infants presenting to the emergency department with lower respiratory tract infection or apnea. *The Pediatric Infectious Disease Journal*, 32(4), 335–340. <https://doi.org/10.1097/inf.0b013e318282603a>
- Kristensen, K., Hjuler, T., Ravn, H., Simões, E. a. F., & Stensballe, L. G. (2012). Chronic diseases, chromosomal abnormalities, and congenital malformations as risk factors for respiratory syncytial virus hospitalization: A Population-Based Cohort study. *Clinical Infectious Diseases*, 54(6), 810–817. <https://doi.org/10.1093/cid/cir928>
- Larrañaga, C., Ampuero, S., Luchsinger, V., Carrión, F., Aguilar, N. V., Morales, P. R., Palomino, M. A., Tapia, L. I., & Avendaño, L. F. (2009). Impaired immune response in severe human lower tract respiratory infection by respiratory syncytial virus. *The Pediatric Infectious Disease Journal*, 28(10), 867–873. <https://doi.org/10.1097/inf.0b013e3181a3ea71>
- Martin, E. T., Kuypers, J., Heugel, J., & Englund, J. A. (2008). Clinical disease and viral load in children infected with respiratory syncytial virus or human metapneumovirus. *Diagnostic Microbiology and Infectious Disease*, 62(4), 382–388. <https://doi.org/10.1016/j.diagmicrobio.2008.08.002>
- Mazur, N. I., Martínón-Torres, F., Baraldi, E., Fauroux, B., Greenough, A., Heikkinen, T., Manzoni, P., Mejías, A., Nair, H., Papadopoulos, N. G., Polack, F. P., Ramilo, O., Sharland, M., Stein, R. T., Madhi, S. A., & Bont, L. (2015). Lower respiratory tract infection caused by respiratory syncytial virus: current management and new therapeutics. *The Lancet. Respiratory Medicine (Print)*, 3(11), 888–900. [https://doi.org/10.1016/s2213-2600\(15\)00255-6](https://doi.org/10.1016/s2213-2600(15)00255-6)
- Mehta, R., Scheffler, M., Tapia, L. I., Aideyan, L., Patel, K., Jewell, A. M., Avadhanula, V., Mei, M., Garofalo, R. P., & Piedra, P. A. (2014). Lactate dehydrogenase and caspase activity in nasopharyngeal secretions are

- predictors of bronchiolitis severity. *Influenza and Other Respiratory Viruses*, 8(6), 617–625.
<https://doi.org/10.1111/irv.12276>
- Meissner, H. C. (2016). Viral bronchiolitis in children. *The New England Journal of Medicine (Print)*, 374(1), 62–72. <https://doi.org/10.1056/nejmra1413456>.
- Mejías, A., & Ramilo, O. (2015). New options in the treatment of respiratory syncytial virus disease. *Journal of Infection*, 71, S80–S87. <https://doi.org/10.1016/j.jinf.2015.04.025>
- Mella, C., Suarez-Arrabal, M. C., Lopez, S. M. C., Stephens, J., Fernández, S., Hall, M. W., Ramilo, O., & Mejías, A. (2012). Innate Immune Dysfunction is Associated with Enhanced Disease Severity In Infants with Severe Respiratory Syncytial Virus Bronchiolitis. *The Journal of Infectious Diseases*, 207(4), 564–573. <https://doi.org/10.1093/infdis/jis721>
- Oey, A., McClure, M. W., Symons, J., Chanda, S., Fry, J., Smith, P. F., Luciani, K., Fayon, M., Chokephaibulkit, K., Uppala, R., Bernatoniene, J., Furuno, K., Stanley, T., Huntjens, D., & Witek, J. (2023). Lumicitabine, an orally administered nucleoside analog, in infants hospitalized with respiratory syncytial virus (RSV) infection: Safety, efficacy, and pharmacokinetic results. *PloS One*, 18(7), e0288271. <https://doi.org/10.1371/journal.pone.0288271>
- Ogra, P. L. (2004). Respiratory syncytial virus: The virus, the disease and the immune response. *Paediatric Respiratory Reviews*, 5, S119–S126. [https://doi.org/10.1016/s1526-0542\(04\)90023-1](https://doi.org/10.1016/s1526-0542(04)90023-1)
- Openshaw, P. (2002). Potential therapeutic implications of new insights into respiratory syncytial virus disease. *Respiratory Research*, 3(S1). <https://doi.org/10.1186/rr184>
- Osterweil, D., & Norman, D. C. (1990). An outbreak of an Influenza-Like illness in a nursing home. *Journal of the American Geriatrics Society*, 38(6), 659–662. <https://doi.org/10.1111/j.1532-5415.1990.tb01425.x>
- Palivizumab, a humanized respiratory syncytial virus monoclonal antibody, reduces hospitalization from respiratory syncytial virus infection in high-risk infants. (1998). *Pediatrics*, 102(3), 531–537. <https://doi.org/10.1542/peds.102.3.531>
- Saleeby, C. M. E., Bush, A. J., Harrison, L. M., Aitken, J. A., & DeVincenzo, J. P. (2011). Respiratory syncytial virus load, viral dynamics, and disease severity in previously healthy naturally infected children. *The Journal of Infectious Diseases*, 204(7), 996–1002. <https://doi.org/10.1093/infdis/jir494>
- Shay, D. K., Holman, R. C., Newman, R. D., Liu, L. L., Stout, J., & Anderson, L. J. (1999). Bronchiolitis-Associated hospitalizations among US children, 1980-1996. *JAMA*, 282(15), 1440. <https://doi.org/10.1001/jama.282.15.1440>
- Sheeran, P., Jafri, H. S., Carubelli, C. M., Saavedra, J., Johnson, C., Krisher, K., Sánchez, P. J., & Ramilo, O. (1999). Elevated cytokine concentrations in the nasopharyngeal and tracheal secretions of children with respiratory syncytial virus disease. *The Pediatric Infectious Disease Journal*, 18(2), 115–122. <https://doi.org/10.1097/00006454-199902000-00007>
- Sigurs, N. (2001). Epidemiologic and clinical evidence of a respiratory Syncytial Virus–Reactive Airway Disease link. *American Journal of Respiratory and Critical Care Medicine*, 163(supplement_1), S2–S6. https://doi.org/10.1164/ajrccm.163.supplement_1.2011109
- Staat, M. A. (2002). Respiratory syncytial virus infections in children. *Seminars in Respiratory Infections*, 17(1), 15–20. <https://doi.org/10.1053/srin.2002.31688>
- Sugawara, M., Czaplicki, J., Ferrage, J., Haeuw, J., Power, U. F., Corvaia, A. N., Nguyen, T. N., Beck, A., & Milton, A. (2002). Structure–antigenicity relationship studies of the central conserved region of human respiratory syncytial virus protein G. *Journal of Peptide Research*, 60(5), 271–282. <https://doi.org/10.1034/j.1399-3011.2002.21027.x>
- Thompson, W., Shay, D. K., Weintraub, E., Brammer, L., Cox, N. J., Anderson, L. J., & Fukuda, K. (2003). Mortality associated with influenza and respiratory syncytial virus in the United States. *JAMA*, 289(2), 179. <https://doi.org/10.1001/jama.289.2.179>

- Welliver, R. C., Wong, D., Sun, M., Middleton, E., Vaughan, R. S., & Ogra, P. L. (1981). The Development of Respiratory Syncytial Virus-Specific IgE and the Release of Histamine in Nasopharyngeal Secretions after Infection. *The New England Journal of Medicine*, 305(15), 841–846. <https://doi.org/10.1056/nejm198110083051501>
- Wright, P. F., Gruber, W. C., Peters, M. R., Reed, G. W., Zhu, Y., Robinson, F. W., Coleman-Dockery, S. D., & Graham, B. S. (2002). Illness Severity, Viral Shedding, and Antibody Responses in Infants Hospitalized with Bronchiolitis Caused by Respiratory Syncytial Virus. *The Journal of Infectious Diseases*, 185(8), 1011–1018. <https://doi.org/10.1086/339822>
- Wright, P. F., Gruber, W. C., Peters, M. R., Reed, G. W., Zhu, Y., Robinson, F. W., Coleman-Dockery, S. D., & Graham, B. S. (2002b). Illness Severity, Viral Shedding, and Antibody Responses in Infants Hospitalized with Bronchiolitis Caused by Respiratory Syncytial Virus. *The Journal of Infectious Diseases*, 185(8), 1011–1018. <https://doi.org/10.1086/339822>
- Xing, Y., & Proesmans, M. (2019). New therapies for acute RSV infections: where are we? *European Journal of Pediatrics*, 178(2), 131–138. <https://doi.org/10.1007/s00431-018-03310-7>
- Zamora, M. R., Budev, M., Rolfe, M. W., Gottlieb, J., Humar, A., DeVincenzo, J. P., Vaishnav, A., Cehelsky, J., Albert, G., Nochur, S. V., Gollob, J., & Glanville, A. R. (2011). RNA Interference Therapy in Lung Transplant Patients Infected with Respiratory Syncytial Virus. *American Journal of Respiratory and Critical Care Medicine*, 183(4), 531–538. <https://doi.org/10.1164/rccm.201003-0422oc>